A New Manufacturing Process to Remove Thrombogenic Factors (II, VII, IX, X, and XI) from Intravenous Immunoglobulin Gamma Preparations

INTRODUCTION

- The risk of thrombi formation and thromboembolic events is increased due to high concentrations of coagulation factors II, VII, IX, X, and particularly XIa remaining in intravenous immunoglobulin (IVIG) preparations as a result of imperfect removal during the manufacturing process.^{1,2}
- IVIG preparation via ethanol precipitation, a process developed in the 1940s, effectively removes coagulation factors II, VII, IX, and X.^{3,4}
- The presence of factor XIa (FXIa), however, still poses a risk of activating the coagulation cascade in the blood of patients receiving IVIG.
- It is difficult to separate FXIa from IgG via ethanol precipitation alone due to the high isoelectric point (pl 8.9-9.1) of FXIa.⁵
- An additional purification process is therefore required to remove residual FXIa in IVIG preparations.

STUDY OBJECTIVES

- To develop a new process for reducing the content of FXIa in IVIG preparations to levels below detectable limits
- To evaluate the robustness of FXIa removal in the new manufacturing process via a spiking study

COAGULATION CASCADE

Figure 1. The Coagulation Cascade





Procoagulant activities were reduced to low levels as determined by:

- < 1.56 mIU/mL
- Chromogenic FXIa Assay < 0.16 mIU/mL
- Non-activated Partial Thromboplastin Time (NaPTT) > 250 seconds
- FXI/FXIa ELISA < 0.31 ng/mL

IVIG PURIFICATION PROCESS AND QUANTITATION OF FXIa

The chromatographic cation exchange (CEX) process utilizes a new ceramic-based resin that binds IgG with 1.5 times higher capacity compared to commonly used resins in protein purification.

Procoagulant activity of the intermediate and final (GC5101B) products was evaluated using:

- Thrombin generation assay (TGA)
- Chromogenic FXIa assay
- Non-activated partial thromboplastin time (NaPTT)
- FXI/FXIa ELISA
- Western blot

Adjusting the pH of the elution buffer to 4.5 and conducting ultra/diafiltration resulted in maximal inhibition of IgG polymerization – thus establishing a stable manufacturing process for GC5101B.

RESULTS

Thrombin Generation Assay (TGA)

Figure 3. FXIa Remaining after Cold Ethanol Fractionation

FXIa content for each process was calculated using the step residual ratio.

- **1.** Cryo-poor plasma (statistical results of 3 batches, mean ± SD): 100%
- **2.** Fraction I + II + III paste: 74.4 ± 3.7
- **3.** Fraction I + III supernatant: 5.8 ± 2.7
- **4.** Fraction II paste: 5.2 ± 2.1. Results from 3 consecutive batches are shown; all samples measured at least in duplicate.



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RESULTS

Table 1. Results of Spiking Study

Evaluation of the reduction of FXIa contents and activity by the spiking test in cation exchange chromatography (CM Hyper D).

Process step	Pre-CEX w/o spiked		Pre-CEX w/ spiked ^d			
	ELISA ^a (µg/L)	TGA ^b (IU/L)	ELISA (µg/L)	TGA (IU/L)	ELISA (µg/L)	TGA (IU/L)
Pre-CEX	123.6	42.2	4020.1	72837.4	21002.2	447861.4
Post-CEX	<0.31	<1.56	<0.31	<1.56	2.93	<1.56
lgG recovery (%)	97.1		95.3		99.4	
Residual ratio (%)	N.C. ^c	N.C.	N.C.	N.C.	0.01	N.C.
All samples were measured at least in duplicate. ^a ELISA: Enzyme-linked immunosorbent assay			32.5X more normal	e FXIa than plasma	170X more normal	e FXIa than plasma

^b TGA: Thrombin generation assay

^c N.C.: Not calculated

^d These concentrations are 32.5 times and 170 times higher than those in normal specimens.



FXIa reduced to below detection limits



RESULTS

Figure 4. Removal of Coagulation Factors by Cold Ethanol Fractionation and CEX Chromatography



A. Image of the western blot with control (non-reduced condition, 4-20% Tris-glycine gel, upper arrow: IgG monomer, 150 kDa; lower arrow: IgG polymer) Loading amount is 5.0 mg of IgG and concentration of FXI/ FXIa calculated by ELISA

- **4.** Fraction II paste (13.0 ng/mL)
- **5.** Pre-CEX chromatography (10.3 ng/mL)
- **6.** Post-CEX chromatography (not detected)



B. Band density of FXI/FXIa calculated using Prizm ver. 6.0

DISCUSSION

- Coagulation factors II, VII, IX, and X were removed via cold ethanol precipitation and FXIa was successfully removed to undetectable levels using CEX chromatography.
- The robustness of this process was evaluated through a spiking study. With the spiking assay, we showed that the high binding capacity resin (> 100.0 g/L) further increased the production efficiency.
- The spiking study including cation exchange chromatography was conducted using 32.5 and 170 times the concentration of FXIa as that present in normal specimens, and the results were analyzed using ELISA and TGA.
- The results demonstrate the ability of CEX chromatography to effectively remove procoagulant activities to below the detection limit (except by NaPTT).
- A reduced risk of thromboembolic events may result from this new IVIG fractionation process.

CONCLUSION

- The use of our manufacturing process may improve the safety of IVIG by reducing FXIa levels to below the detection limit, potentially minimizing the risk of thromboembolic events associated with the use of IVIG products.
- Our results are difficult to directly compare with previous studies, as we have used different test and standard materials to quantitate FXIa activity.
- Additional in vivo assays (such as the Wessler test) would be necessary to demonstrate the safety of the preparations more precisely.

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